In the Specification:

On page 6, at line 34, please delete the following paragraph:

~~Reagents suitable for use in labelling these markers can be found in Table 4.~~

On page 22, the first full paragraph bridging page 23 should be amended to read:

Telomerase Repeat Amplification Protocol (TRAP) Assay

Telomerase activity was measured by a modified non-radioactive TRAP protocol essentially as described by Fong *et al* (1997). Telomerase cell extracts were prepared by the method of Kim *et al,* (1994), with minor modifications. Populations of sorted or cultured cells were lysed in ice-cold CHAPS extraction buffer (0.5% 3[(3-cholamidopropyl)-dimethyl-ammonio]-1-propanesulfonate], 50 mM Tris-HCI, pH 7.4, 5 mM $MgCl_2$, 5 mM EGTA, 25 mM 2-mercaptoethanol, 1 ng/ml leupeptin, and 50% glycerol in DEPC-treated water), at a concentration of 1000 cells/µl, incubated on ice for 30 minutes and centrifuged at 16000 xg for 20 minutes at 4°C, the supernatant recovered and stored at -80°C until required. Detection of telomerase activity was performed in a two-step process as previously described (Fong *et al*, 1997). Briefly, to 2µl of cell extract, 16.5 µl of TRAP reaction buffer (20 mM Tris-HCI, pH8.2, 1.5mM $MgCl_2$, 63 mM KCl, 0.05%Tween-20, 1 mM EGTA), 100 ng of each of TS primer (5'-AATCCGTCGAGCAGAGTT-3', SEQ ID NO: 1), and CX-ext primer (5'-GTGCCCTTCCCTTACCCTTACCC TAA-3', SEQ ID NO: 2) , 0.5 µL dNTPs (10 mM stock) were added, and the reaction mix incubated at 25°C for 30 minutes. Telomerase was subsequently inactivated by heating the reaction to 90°C for 2 minutes, prior to the addition of 5 µl of PCR mixture, containing 3.5 µl of TRAP reaction buffer, 1 µl of CX-ext primer and 2.5 U Taq polymerase. Reaction mixes were covered with mineral oil and placed in a Hybaid thermocycler, and subjected for 34 cycles of 94°C for 30 seconds, 50°C for 30 seconds and 72°C for 45 seconds, with a final extension at 72°C for 2 minutes. To confirm the specificity of the telomerase products, in all cases, a 2 µl aliquot of each CHAPS lysate was subjected to denaturation by heating samples at 100°C for 10 minutes. 25 µl of each reaction was resolved on a non-denaturing 12% polyacryalmide gel, and visualised by staining width SYBR Green fluorescent dye (FMC Bioproducts, OR, USA) as recommended by the manufacturer. The TRAP products were analysed using a fluorescence scanning system (Molecular Dynamics, Sunnyvale, CA, USA).